

## **resDetect™ T7 RNA Polymerase ELISA Kit (Residue Testing)**

**Pack Size: 96 tests**

**Catalog Number: RES-A018**

***IMPORTANT: Please carefully read this manual before performing your experiment.***

***For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedure***

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## Content

<b>【Product Overview】</b> .....	1
<b>【Assay Principle】</b> .....	1
<b>【Precautions】</b> .....	1
<b>【Materials Provided】</b> .....	2
<b>【Storage】</b> .....	2
<b>【Reagents and Consumables / Equipment Required but not Provided】</b> .....	3
<b>【Quick Guide】</b> .....	4
<b>【Reagent Preparation】</b> .....	5
<b>【Assay Procedure】</b> .....	5
<b>【Calculation of Results】</b> .....	9
<b>【Typical Data】</b> .....	10
<b>【Sensitivity】</b> .....	10
<b>【Precision】</b> .....	10
<b>【Recovery】</b> .....	11
<b>【Linearity】</b> .....	11
<b>【Troubleshooting Guide】</b> .....	12

## 【Product Overview】

resDetect™ T7 RNA Polymerase ELISA Kit (Residue Testing) is designed to detect and quantitatively determine T7 RNA Polymerase residues in mRNA drugs. The kit contains T7 RNA Polymerase to ensure accurate assay results and is designed to provide a reliable solution for monitoring and controlling T7 RNA Polymerase residues during the production of mRNA products. It can also be used as a universal detection tool for the quantitative determination of T7 RNA Polymerase.

## 【Assay Principle】

This kit quantifies T7 RNA Polymerase using a sandwich ELISA format. The microplate is pre-coated with an Anti-T7 RNA polymerase Antibody, which captures T7 RNA Polymerase present in standards and samples. After washing, a Biotin-Anti-T7 RNA polymerase Antibody is added to bind the captured T7 RNA Polymerase, forming an Antibody-antigen-biotinylated antibody sandwich complex. After washing, a Streptavidin-HRP is added to the plate. Following additional washes, a substrate is added for color development. The reaction is stopped with stop solution, and the color changes from blue to yellow. Absorbance is measured at 450 nm with a 630 nm reference. The absorbance signal is directly proportional to the T7 RNA Polymerase concentration in the sample.

## 【Precautions】

1. **For research use only.** Not for use in diagnostic or therapeutic procedures.
2. **Use before the expiration date** indicated on the label.
3. Do not mix or interchange reagents from different lots or kits.
4. If a sample OD exceeds the highest standard dilute the sample with 1×Dilution Buffer and retest.
5. Result variability may arise from operator technique, equipment performance, incubation conditions, or storage/handling. Follow the protocol exactly. The kit is designed to reduce some endogenous interference in biological samples, however, not all potential interfering factors can be eliminated.

## 【Materials Provided】

Table 1. Materials Provided

ID	Components	Size (96 T)	Format	Storage	
				Unopened	Opened
RES018-C01	Pre-coated Anti-T7 RNA polymerase Antibody Microplate	1 plate	Solid	2-8°C	2-8°C
RES018-C02	T7 RNA polymerase Standard	100 µL	Liquid	2-8°C	2-8°C
RES018-C03	Biotin-Anti-T7 RNA polymerase Antibody	150 µL	Liquid	2-8°C	2-8°C
RES018-C04	Streptavidin-HRP	50 µL	Liquid	2-8°C, Protected from light	2-8°C, Protected from light
RES018-C05	20×Washing Buffer	50 mL	Liquid	2-8°C	2-8°C
RES018-C06	Biotin-Antibody and Streptavidin-HRP Dilution Buffer	50 mL	Liquid	2-8°C	2-8°C
RES018-C07	Standard and Sample Dilution Buffer (5×)	30 mL	Liquid	2-8°C	2-8°C
RES018-C08	Substrate Solution	12 mL	Liquid	2-8°C, Protected from light	2-8°C, Protected from light
RES018-C09	Stop Solution	7 mL	Liquid	2-8°C	2-8°C

Note: Briefly centrifuge the Streptavidin-HRP before use to bring the contents to the bottom of the tube.

## 【Storage】

1. Store the unopened kit at 2-8°C upon receipt.
2. Locate the expiration date on the outer packaging and do not use reagents beyond their expiration date.

**【Reagents and Consumables / Equipment Required but not Provided】**

**Table 2. Reagents and Consumables / Equipment Required but not Provided**

Items	Specification
Deionized or distilled water	/
Single- or multi-channel micropipettes	Calibrated
Low-retention pipette tips	10 $\mu$ L, 100 $\mu$ L, 300 $\mu$ L, 1000 $\mu$ L
Reagent bottle	500 mL
Centrifuge tubes	1.5 mL, 10 mL
Vortex mixer	/
Timer	/
Incubator	37°C
Microplate reader	Single- or dual- wavelength microplate reader capable of measuring absorbance at 450 nm with a 630 nm reference in 96-well microplates.

**【Quick Guide】**

**Step 1 Preparation of the Standard Curve**

Prepare the serial dilutions:

Tubes/Solution Code	Standard Stock Solution	Std.-0	Std.-1	Std.-2	Std.-3	Std.-4	Std.-5	Std.-6	Std.-7
Procedure	10 $\mu$ L	15 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L
Solution Conc.	200 $\mu$ g/mL	2000 ng/mL	50 ng/mL	25 ng/mL	12.5 ng/mL	6.25 ng/mL	3.13 ng/mL	1.56 ng/mL	0.78 ng/mL
Dilution Buffer Vol.		990 $\mu$ L	585 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L	300 $\mu$ L

**Step 2 Add standards, samples, and blank controls**



Addition: Add 100  $\mu$ L of standards, samples, and blank to the designated wells.



Incubation: Seal the plate and incubate at room temperature for 1.0 h.

**Step 3 Add Detection Antibody**



Washing (manual or automated): Discard the liquid. Add 300  $\mu$ L of wash buffer, wash 3 times, and tap dry.



Addition: Add 100  $\mu$ L to each well.



Incubation: Seal the plate and incubate at room temperature for 1.0 h.

**Step 4 Add Streptavidin-HRP**



Washing (manual or automated): Discard the liquid. Add 300  $\mu$ L of wash buffer, wash 3 times, and tap dry.




Addition: Add 100  $\mu$ L to each well.

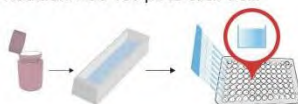


Incubation: Seal the plate and incubate at room temperature for 1.0 h.


**Step 5 Add Substrate Solution**



Washing (manual or automated): Discard the liquid. Add 300  $\mu$ L of wash buffer, wash 3 times, and tap dry.



Addition: Add 100  $\mu$ L to each well.



Incubation: Seal the plate and incubate at room temperature for 20 min, protected from light.

**Step 6 Add Stop Solution and Data Recording**



Addition: Add 50  $\mu$ L of Stop Solution.  
Note: The color will change from blue to yellow.



Data Recording: Measure the absorbance at 450 nm with a 630 nm reference within 5 min after adding the stop solution.

## 【Reagent Preparation】

Bring all reagents to room temperature (20-25°C) before use. If crystals are present in the solution, allow the reagents to equilibrate until the crystals are completely dissolved. If needed, incubate at 37°C for 10-15 minutes to facilitate dissolution.

## 【Assay Procedure】

### 1. Preparation of Working Solution

#### 1.1 Preparation of 1×Washing Buffer

Dilute 50 mL of 20×Washing Buffer with ultrapure water or deionized water to a final volume of 1000 mL and mix gently.

#### 1.2 Preparation of 1×Standard and Sample Dilution Buffer

Dilute 30 mL of 5×Dilution Buffer with 1×Washing Buffer to a final volume of 150 mL and mix gently.

#### 1.3 Preparation of Biotin-Anti-T7 RNA polymerase Antibody working solution

Dilute the Biotin-Anti-T7 RNA polymerase Antibody at 1:200 using Biotin-Antibody and Streptavidin-HRP Dilution Buffer. Prepare fresh before use.

#### 1.4 Preparation of Streptavidin-HRP working solution

Dilute the Streptavidin-HRP at 1:2000 using Biotin-Antibody and Streptavidin-HRP Dilution Buffer. Prepare fresh before use, protected from light.

### 2. Preparation of the Standard Curve

The concentration of the T7 RNA polymerase Standard (RES018-C02) is 200 µg/mL.

#### 2.1 Prepare Std.-0

Dilute 10 µL of the T7 RNA polymerase Standard into 990 µL of 1×Standard and Sample Dilution Buffer. Mix gently and thoroughly.

#### 2.2 Prepare the highest standard (Std.-1, 50 ng/mL)

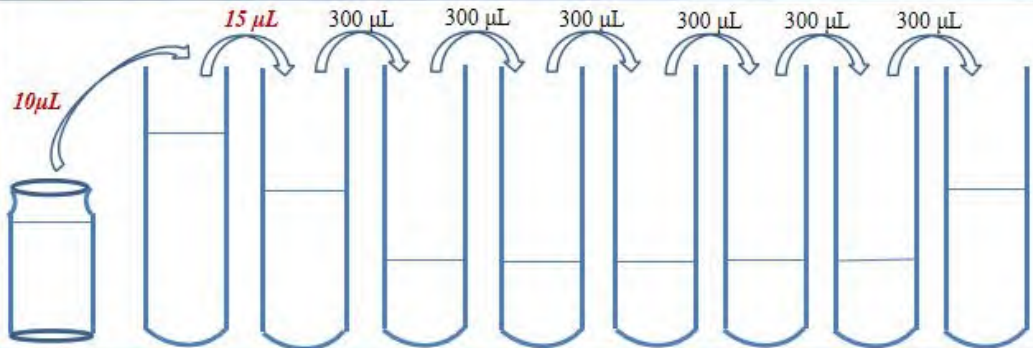
Dilute 15 µL of Std.-0 into 585 µL of 1×Standard and Sample Dilution Buffer. Mix gently and thoroughly.

#### 2.3 Prepare the Serial Dilutions

Add 300 µL of 1×Standard and Sample Dilution Buffer to each of the remaining centrifuge tubes. Perform 2-fold serial dilutions starting from Std.-1 to generate the standard curve. Mix thoroughly at each dilution step. 1×Standard and Sample Dilution Buffer serves as the zero standard (Blank). The dilution procedure is shown in Figure 1.

*Note: For residual analysis, it is recommended to generate the standard curve using standards from the same manufacturer and production lot as the target analyte to minimize measurement variability caused by differences in quantification methods or other supplier-related factors.*

**Figure 1. Preparation of 2-Fold Serial Dilutions of the T7 RNA Polymerase Standard**

Tubes/ Solution Code	Standard stock solution	Std.-0	Std.-1	Std.-2	Std.-3	Std.-4	Std.-5	Std.-6	Std.-7
Procedure									
Solution Conc.	200 µg/mL	2000 ng/mL	50 ng/mL	25 ng/mL	12.5 ng/mL	6.25 ng/mL	3.13 ng/mL	1.56 ng/mL	0.78 ng/mL
Dilution Buffer Vol.		990 µL	585 µL	300 µL	300 µL	300 µL	300 µL	300 µL	300 µL

### 3. Preparation of Samples (For reference)

Spike recovery experiments are a critical component of ELISA analysis. They are designed to assess potential interference from the sample matrix on target analyte detection and verify the reliability of the assay method. The core principle of this approach is to spike a known quantity of standard material into samples with a pre-determined concentration; recovery is then calculated by comparing the measured signal increment to the theoretical amount of standard added.

#### 3.1 Pretreatment of Test Samples

Serially dilute the test samples with 1×Standard and Sample Dilution Buffer to ensure that the analyte concentration falls within the detection range of the standard curve and to reduce potential matrix interference. The dilution procedure is shown in Table 3.

**Table 3. Preparation Method**

Sample ID	Volume(µL)	1×Standard and Sample Dilution Buffer(µL)	Total volume(µL)	Final Dilution Ratio	Diluted Sample ID
Test sample	150	150	300	2	Test sample-1
Test sample-1	150	150	300	4	Test sample-2

*Note: 1) The required dilution factor may vary depending on the sample matrix and should be*

*optimized for each sample type.*

*2) Perform serial dilutions carefully to ensure accuracy. It is recommended that the dilution factor for any single step does not exceed 10-fold, as excessive dilution may affect assay accuracy.*

### 3.2 Preparation of Spiking Working Solution

Based on the expected sample concentrations, dilute the standard to an appropriate concentration for the spiking working solution using 1×Standard and Sample Dilution Buffer. Prepare the spiking solution freshly before use.

*Note: The final concentration of analyte in the spiked samples should fall within the linear detection range of the assay to avoid inaccurate results caused by values outside the assay range.*

### 3.3 Preparation of Spiked Samples

Mix the test samples prepared in Step 3.1 with either the spike working solution or 1×Dilution Buffer at a 1:1 (v/v) ratio to prepare spiked and unspiked samples, respectively. Mix thoroughly. The dilution scheme is shown in Table 4.

*Note: To minimize procedural variation, ensure that the sample volume, diluent or spike volume, mixing method, incubation time, and incubation temperature are identical for both spiked and unspiked samples.*

**Table 4. Preparation Method**

Sample ID	Group	Volume(μL)	1×Standard and Sample Dilution Buffer(μL)	Spiking working solution(μL)	Total volume(μL)	Final Dilution Ratio
Test sample	Unspiked	150	150	0	300	2
Test sample	Spiked	150	0	150	300	2
Test sample-1	Unspiked	150	150	0	300	4
Test sample-1	Spiked	150	0	150	300	4
Test sample-2	Unspiked	150	150	0	300	8
Test sample-2	Spiked	150	0	150	300	8

## 4. Addition of Samples

Add 100 μL serially diluted standards, test samples and spiked samples to the appropriate wells. Add 100 μL of 1×Standard and Sample Dilution Buffer to the Blank control wells.

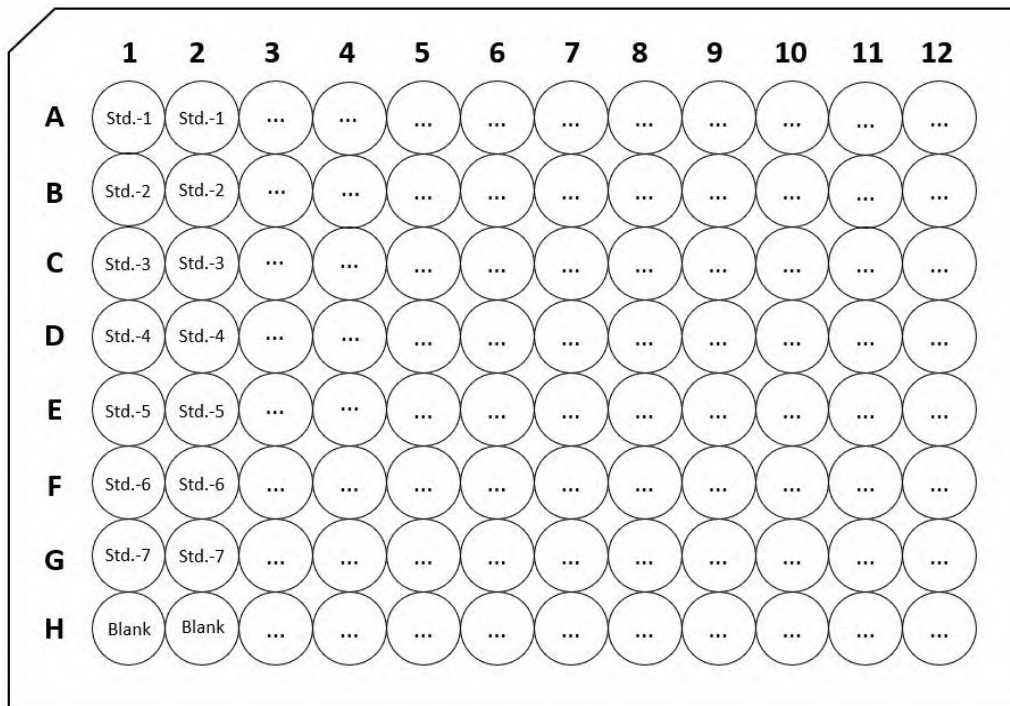
*Note: 1) It is recommended to run all standards and samples in duplicate wells.*

*2) The Blank control contains 1×Standard and Sample Dilution Buffer only.*

*3) It is recommended to use the example plate layout shown in Figure 2 to record the positions of*

standards and samples.

**Figure 2. Recommended Plate Layout for Standards and Samples**



**5. Incubation**

Seal the plate with microplate sealing film. Incubate at room temperature for 1.0 hour.

**6. Washing**

Carefully remove the plate sealer and discard the liquid from the wells. Add 300 µL of 1×Washing Buffer to each well and soak for 10 seconds. Aspirate or decant the buffer. Repeat for a total of three washes. After the final wash, invert the plate and blot dry on absorbent paper.

**7. Addition of Detection Antibody**

Add 100 µL Biotin-Anti-T7 RNA polymerase Antibody working solution (1:200 dilution) to each well. The working solution should be prepared fresh before use.

**8. Incubation**

Seal the plate with microplate sealing film. Incubate at room temperature for 1.0 hour.

**9. Washing**

Repeat step 6.

**10. Addition of Streptavidin-HRP**

Add 100 µL Streptavidin-HRP working solution (1:2000 dilution) to each well. Prepare the working solution fresh before use.

## 11. Incubation

Seal the plate with microplate sealing film. Incubate at room temperature for 1.0 hour, protected from light.

## 12. Washing

Repeat step 6.

## 13. Substrate Reaction

Add 100 µL Substrate Solution to each well. Seal the plate and incubate at room temperature for 20 minutes, protected from light.

## 14. Reaction Termination

Add 50 µL of Stop Solution to each well. Gently tap the plate to ensure thorough mixing.

*Note: The color in the wells will change from blue to yellow.*

## 15. Data Recording

Measure the absorbance at 450 nm with a 630 nm reference within 5 minutes after adding the stop solution.

*Note: Subtracting the OD<sub>630nm</sub> value from the OD<sub>450nm</sub> value helps reduce background interference.*

### 【Calculation of Results】

1. Calculate the mean absorbance of duplicate wells for each standard and sample. Subtract the absorbance of the zero standard (Blank) from each value (OD<sub>450 nm</sub>-OD<sub>630 nm</sub>-Blank).
2. Generate the standard curve by plotting the standard concentrations on the x-axis and the corrected absorbance values on the y-axis. Fit the curve using a four-parameter logistic (4-PL) model. The coefficient of determination (R<sup>2</sup>) should be ≥ 0.9900.
3. Determine the concentrations of the samples and spiked samples from the standard curve. Multiply the calculated concentrations by the appropriate dilution factors.
4. Calculation of spiked recovery (%)

$$\text{Recovery Rate (\%)} = \frac{\text{Concentration Spiked Sample} - \text{Concentration Unspiked Sample}}{\text{Concentration Control Spike}} \times 100\%$$

If the spike recovery falls outside the range of 80%-120%, this may indicate matrix interference affecting the ELISA assay. In such cases, optimize sample pre-treatment (e.g., additional dilution) before retesting.

## 5. Detection Range

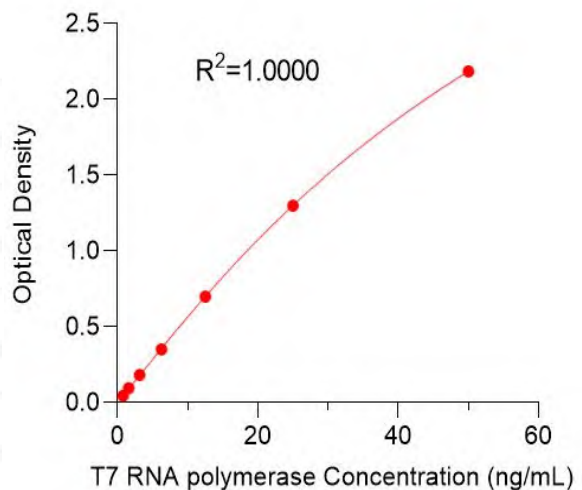
The assay detection range is 0.78 ng/mL-50 ng/mL. Samples with concentrations above the upper limit of the calibration curve should be reported as >50 ng/mL or diluted appropriately and

reanalyzed to fall within the linear range. Samples with concentration below the lower limit of calibration curve should be reported as <0.78 ng/mL.

**【Typical Data】**

A standard curve must be generated for each microplate in every experiment. Absolute OD values may vary depending on the laboratory, operator, and equipment used. The example data provided below are for reference only.

Standard (ng/mL)	O.D.-1	O.D.-2	Average	Corrected
50	2.213	2.293	2.253	2.186
25	1.338	1.400	1.369	1.302
12.5	0.771	0.766	0.769	0.701
6.25	0.416	0.425	0.421	0.353
3.13	0.253	0.250	0.252	0.184
1.56	0.167	0.161	0.164	0.097
0.78	0.110	0.117	0.114	0.046
0	0.069	0.066	0.068	/



**【Sensitivity】**

The minimum detectable concentration (MDC) of T7 RNA polymerase is 0.161 ng/mL. The MDC was determined by calculating the mean optical density (OD) of 20 zero-standard replicates, adding two standard deviations (mean+2SD), and converting the resulting OD value to concentration using the standard curve.

**【Precision】**

**1. Intra-assay Precision**

Three samples of known concentration were tested twenty replicates within a single plate to evaluate intra-assay precision.

**2. Inter-assay Precision**

Three samples of known concentration were tested in three independent assays to evaluate inter-assay precision.

Sample	Intra-assay Precision			Inter-assay Precision		
	1	2	3	1	2	3
n	20	20	20	3	3	3
Mean (ng/mL)	36.606	12.845	1.779	37.457	13.009	1.801
SD	2.255	0.383	0.149	0.805	0.213	0.023
CV (%)	6.2	3.0	8.4	2.1	1.6	1.3

### 【Recovery】

Three samples at different concentration levels were evaluated to determine the spike recovery of the assay.

Sample(n=5)	Average Recovery (%)	Range (%)
High	88.3	82.9-93.6
Middle	92.9	86.5-100.2
Low	93.3	81.4-107.7

### 【Linearity】

To assess the assay linearity, a high-concentration standard was spiked into different dilution matrices and serially diluted to generate concentrations within the dynamic range of the assay.

		Cell culture medium (DMEM)	Cell culture medium (1640)
1:2	Average Recovery (%)	97.4	100.5
	Range (%)	94.1-102.3	98.5-102.7
1:4	Average Recovery (%)	97.9	97.0
	Range (%)	94.8-102.1	88.3-103.3
1:8	Average Recovery (%)	103.8	103.0
	Range (%)	86.2-110.8	98.6-109.2
1:16	Average Recovery (%)	103.5	95.2
	Range (%)	96.5-107.4	87.4-103.1

## 【Troubleshooting Guide】

Problem	Cause	Solution
Low signal	a. Kit components were not equilibrated to room temperature before use; b. Insufficient reconstitution time for lyophilized components.	a. Remove the kit from 2-8 °C storage in advance and allow all reagents to fully equilibrate to room temperature before starting the assay; b. After adding reconstitution buffer, allow lyophilized components to stand for at least 15 minutes and mix gently before use.
Poor assay reproducibility	a. Improper storage of reagents after opening; b. In consistent timing during sample dilution or pipetting.	a. Store reagents strictly according to the instructions in this manual; b. Plan the experimental workflow in advance and properly schedule assay timing.
High background	a. Insufficient washing; b. Excessive incubation temperature or prolonged substrate development time; c. Reagent contamination.	a. Increase soak time during wash steps and ensure the plate is thoroughly blotted dry after final wash; b. Strictly follow the operating procedures described in the manual; c. Use clean reagents and consumables, and maintain a clean experimental environment.
Edge effects	Uneven temperature distribution across the plate.	Ensure uniform incubation temperature and avoid stacking plates during incubation.
Good standard curve but no detectable signal in samples	a. Interfering substances present in the samples; b. Target analyte concentration below the assay detection.	a. Optimize sample dilution to minimize matrix interference; b. Use a higher-sensitivity assay if lower analyte levels are expected.